

Gene therapy and nasopharyngeal carcinoma*

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Summary

In 2003, a non-replicating adenoviral gene therapy product received the world's first government licence for the treatment of head and neck cancer. Two years later approval was granted to a replication-selective adenovirus for the treatment of nasopharyngeal carcinoma in combination with chemotherapy.

This review introduces the reader to gene therapy as an emerging treatment modality, and outlines its application to the management of nasopharyngeal carcinoma by examining recent pre-clinical and clinical research.

Key words: gene therapy, nasopharyngeal carcinoma, head and neck cancer

Introduction

Nasopharyngeal carcinoma (NPC) has a unique geographical and ethnic predisposition with high incidences in South East Asia (up to 30 per 100,000),⁽¹⁾ North Africa, and the Arctic⁽²⁾. Outside endemic areas the tumour is rare, with an incidence of less than 1 per 100,000⁽³⁾. Interestingly emigrants from endemic areas remain at high risk of developing NPC, which diminishes with successive generations;⁽⁴⁾ suggesting both environmental and genetic aetiological influences. Three well-defined factors for NPC have now been established: genetic susceptibility, early exposure to chemical carcinogens (especially Southern Chinese salted fish), and latent Epstein-Barr virus (EBV) infection⁽⁵⁾. NPC is traditionally divided into three histological subtypes: Type I Keratinising (squamous cell carcinomas), Type II Non-keratinising and Type III Undifferentiated⁽⁶⁾. However, this classification has subsequently been simplified into grade 1 squamous cell carcinomas and grade 2 undifferentiated carcinomas⁽⁷⁾. This latter classification correlates well with endemic area patient origin and EBV status - showing strong association with grade 2 cancers⁽⁸⁾. NPC is generally radiosensitive, and consequently the primary treatment modality is radiotherapy. Five year survival of patients with T1/T2 lesions (Ho classification⁽⁹⁾) range from 75% to 90%,

falling to 50%-75% for T3/4 lesions⁽¹⁰⁾. Intensity modulated radiotherapy (IMRT) has improved local control rates to more than 90%⁽¹¹⁾; as well as reducing toxicity to vital surrounding structures, with resultant improved side effects and quality of life outcomes⁽¹²⁾. Chemotherapy is a radiosensitiser, however its role, and in particular the timing of its delivery in relation to radiotherapy, has not been comprehensively established. The most convincing evidence is for concurrent chemoradiotherapy⁽¹³⁾, although some promising results have been shown for neo-adjuvant chemotherapy^(14,15). The role of surgery, other than in diagnosis through biopsy, is in salvage for local and regional failure. Novel therapies, including gene therapy, have yet to be established in the management of patients with recurrent, residual or metastatic disease following treatment with conventional modalities.

Gene therapy

Gene therapy has been defined as 'the deliberate introduction of genetic material into patient's cells in order to treat or prevent a disease⁽¹⁶⁾'. Gene therapy was initially applied to inherited diseases with single gene mutations (monogenic); such as severe combined immunodeficiency disease (SCID), cystic fibrosis and haemophilia. However, permanent corrective gene

expression in such illnesses has been largely unsuccessful. This, together with the development of leukaemia in children with SCID and the death of a patient with ornithine transcarbamylase deficiency following viral gene therapy, has highlighted the potential risks of such treatments to researchers⁽¹⁷⁾. With this in mind, new applications for gene therapy, such as cancer and cardiovascular disease, were considered; where transient gene delivery would be more likely to be achieved, and provide a safe and clinically significant effect⁽¹⁸⁾.

Cancer is a genetic disease, with normal cells undergoing multiple mutations, as part of an 'oncoevolutionary process,' to transform into malignant cells. These changes can be acquired during life, through exposure to carcinogens, or inherited. Mutations in the cell behaviour regulating genes (proto-oncogenes and tumour suppressor genes) lead to self-sufficiency in growth signals, insensitivity to anti-growth signals, limitless replicative potential, ability to evade apoptosis, sustained angiogenesis, ability to invade tissues and metastasise - the hallmarks of cancer⁽¹⁹⁾. All of these changes are potential targets for gene therapy and has resulted in cancer gene therapy research dominating the gene therapy research field; with two-thirds of clinical trials for cancer applications⁽²⁰⁾. However success at clinical trial has been disappointing mainly due to poor efficiency of gene transfer rather than a paucity of potential therapeutic genes⁽²¹⁾. Strategies to overcome this have focused on modulating the host immune response to the virus, optimising existing viruses and discovering/creating new viruses, and identifying tumour-associated genes that can improve viral potency.

Gene therapy strategies

there are three main strategies that characterise the type of gene expressed in gene therapy: corrective, cytoreductive and immunomodulatory. Cancer is caused by mutations in oncogenes and tumour suppressor genes (TSG) resulting in uncontrolled cell growth. Corrective gene therapy attempts to block oncogene or replace TSG function, thereby returning cells to normal cell growth. In over 50% of all cancers p53 TSG mutations are implicated, and is thus considered to be a potential magic bullet for corrective cancer gene therapy⁽²²⁾. In 2003, the world's first government licensed gene therapy product featured a recombinant adenovirus expressing p53 (Gendicine; SiBiono Schenzhen, China)⁽²³⁾. Oncogenes can be over-expressed or amplified in cancer; and their action can be blocked therapeutically at either the transcriptional or translational level, through RNA interference and catalytic ribozymes respectively.

Cytoreductive gene therapy aims to directly or indirectly kill cancer cells, rather than correct the underlying genetic defect.

This can be achieved by introducing a transgene that can convert an inactive prodrug to a potent anti-cancer agent, for example the cytosine deaminase gene and 5-fluorocytosine (5-FC), which is converted to the highly toxic 5-fluorouracil (5-FU)⁽²⁴⁾. Other strategies include inhibiting angiogenesis, through transgene expression of angiogenesis inhibitors such as endostatin (E10A)⁽²⁵⁾, or inducing apoptosis, by expressing exogenous p53⁽²³⁾.

The host immune response detects tumour associated antigens (TAA), which allows CD8+ cytotoxic T cells to induce tumour lysis. Cancer cells subvert this immuno-surveillance function by reducing MHC class I expression and producing immune-suppressive cytokines⁽²⁶⁾. Gene therapy can reverse this tumour-induced immuno-tolerance by increasing TAA presentation, upregulating MHC I expression or breaking down the immune suppressive environment through transduction of immune modulatory genes⁽²¹⁾. This has been achieved by using cytokines as transgenes, with resultant enhanced antitumour efficacy; for example adenovirus expressing granulocyte macrophage colony-stimulating factor (GM-CSF) in head and neck cancer animal models⁽²⁷⁾. Furthermore clearance of the viral vector by the host immune response can also be reduced by this approach⁽²⁸⁾.

Vectors for gene therapy

The greatest limitation to cancer gene therapy becoming an effective everyday clinical treatment modality is the accuracy and efficiency of gene transfer to the target cells⁽²¹⁾. The ideal vector should be efficient, safe and tumour selective; preferentially affecting cancer cells and sparing normal cells. Tumour selectivity can be achieved by administering the vector directly into the tumour, or through engineering the vector physically or genetically. The latter strategy is more attractive as this would allow for systemic administration, which would be able to treat inaccessible or multiple lesions. However, to reach its target the vector would have to overcome clearance of circulating virus by complement and reticulo-endothelial cell-based mechanisms, which leads to phagocytosis of viral particles by macrophages or Kupffer cells⁽²⁹⁾.

The two main groups of vectors used in cancer gene therapy research are viral and non-viral vectors. Approximately 70% of clinical trials and the majority of research to date has used viral vectors⁽¹⁸⁾. Non-viral vectors use either physical transfection or particle-mediated systems to introduce foreign DNA into the target cells. They are generally less immunogenic than viral vectors which allows for repeated administration⁽³⁰⁾. Additionally they are cheaper to produce and can carry more DNA. However their major disadvantage is low transfection rates.

Viruses are obligate intracellular pathogens, which require host cell machinery to complete their life cycle. They have evolved over millions of years to be highly effective at infecting, replicating within and generally lysing host cells. Viruses have also developed mechanisms that allow them to subvert the host immune system and thereby increase their infectivity, such as expressing interferon γ decoy receptors. Viral vectors commonly induce mild flu-like illnesses, but have the potential for causing acute life-threatening toxicity. Some 'oncolytic' viruses can also exert an anti-cancer effect directly themselves by selectively replicating and destroying cancer cells. Apart from different viral species, a common classification of viral vectors is whether they have the ability to replicate. Non-replicating viruses were the first vector systems for the delivery of foreign genes and most research to date has utilised such viruses⁽³¹⁾. However, the efficacy of gene transfer of these viruses is poor and for successful cancer treatment every tumour cell would have to be infected.

In contrast, replicating viruses allow more cancer cells to be infected as their progeny can go on to infect more cells and destroy the host cell as a direct consequence of their replication⁽³¹⁾. Replication-selective viruses have the ability to replicate in tumour cells, but not normal cells. Some viruses exhibit inherent tumour selectivity, such as Reovirus and Newcastle virus whereas other viruses can be genetically modified to improve tumour selectivity, such as Adenovirus and vaccinia virus. This can be achieved by inserting tumour-specific promoters, such as prostate-specific antigen (PSA) or deleting genes that are needed for viral replication in normal cells but not tumour cells, such as thymidine kinase.

ADENOVIRUS

The vast majority of NPC gene therapy research, indeed most cancer gene therapy research, has used adenovirus mediated strategies⁽³²⁾. Adenoviruses are non-enveloped DNA viruses; first isolated in 1953 from human adenoids. The most significant advantage of adenovirus over other viral vectors is high *in vivo* transfection efficiency⁽³³⁾. Furthermore, adenovirus is able to deliver therapeutic genes to dividing and non-dividing cells, and can be manufactured to high titres⁽³⁴⁾. Also the virus remains episomal and does not integrate into the host cell genome, reducing the risk of insertional oncogenesis. However, adenoviruses are highly immunogenic and, hence, the risk of toxicity and even mortality with their clinical application is a concern⁽³⁵⁾. Fortunately newer engineered viruses containing minimal viral genes ('gutless') exhibit far less toxicity. Adenoviruses have undergone extensive clinical trials in head and neck cancer, with Gendicine (p53 expressing replication deficient adenovirus)^(36,37) and ONYX-015/H101 (E1B deleted replication selective adenovirus)⁽³⁸⁻⁴⁰⁾ the most prominent constructs with established anti-tumour efficacy.

Therapeutic strategies – genetic targets

EBV

EBV is a ubiquitous herpes virus and consistently detected in NPC⁽⁴¹⁾, with lower titres found in the differentiated compared to the undifferentiated forms. EBV is a latent infection in NPC, with the viral genome maintained as a circular episome that replicates once per cell cycle⁽⁴²⁾. One viral protein, Epstein-Barr Nuclear Antigen 1 (EBNA1), and one region of the viral genome, origin of plasmid replication (oriP) have been identified as being necessary for viral replication⁽⁴³⁾. EBNA1 activates viral DNA replication upon binding to specific sequences, termed the family of repeats (FR), within the oriP region of the EBV genome⁽⁴⁴⁾. Additionally EBNA1 binding to the FR element causes downstream enhancement of gene expression, and this has been used as a gene therapy strategy, by restricting certain gene expression to EBV infected cells.

A plasmid has been constructed with the cytosine deaminase gene cloned downstream of the herpes simplex virus thymidine kinase promoter and FR sequence⁽⁴⁵⁾. In EBV-positive cancer cells, transfection with the cytosine deaminase-containing plasmid in the presence of 5-FC abolished cell growth. This strategy has also been used to mediate expression of BRLF1 (RTA); which is an EBV protein able to regulate a switch in EBV replication from latent to lytic replication with resultant destruction of host cells⁽⁴⁶⁾. Recombinant baculovirus constructed with a cytomegalovirus (CMV) promoter, OriP and EBNA1 gene was able to express RTA, induce EBV lytic replication and cell death; as well as inhibit growth in EBV-positive tumours in nude mice⁽⁴³⁾. Further studies have examined Interferon-gamma (IFN- γ) delivery to NPC via a non-viral gene therapy vector under the control of an oriP-CMV promoter⁽⁴⁷⁾. IFN- γ is a critical anti-viral and anti-tumour cytokine, and expression in this vector resulted in selective anti-proliferative effects on EBV-positive cancer cells *in vitro* and *in vivo*. An adenovirus in which p53 transgene expression was under the transcriptional regulation of oriP, resulted in selective gene expression in EBV-positive cells and apoptosis mediated cytotoxicity⁽⁴⁸⁾. A further EBV positive tumour selectivity strategy employs a Virus-associated I (VAI) deleted adenovirus, which showed selective replication in EBV positive tumour cells and superior anti-tumour potency *in vivo*⁽⁴⁹⁾. VAI RNAs are required for translation of adenoviral mRNA, and hence VAI deficient adenoviruses require a substitute source for viral replication. EBV-encoded small RNA1 is expressed in most EBV-associated human tumours and can act to replace the lost VAI RNAs of VAI deleted adenovirus. Another interesting transgene that has been investigated in an adenovirus backbone with oriP FR enhancer elements is Bim_s; which is a pro-apoptotic Bcl-2 family member⁽⁵⁰⁾. The proportion of apoptotic cells following Bim_s expression was significantly increased and tumour regression was seen in mouse xenografts in combination with radiotherapy. The death receptor Fas and

its ligand FasL are extensively expressed in NPC⁽⁵¹⁾, suggesting the Fas-mediated apoptotic pathway is intact. Adenovirus expressed FasL (oriP promoter) shows evidence of inducing apoptosis in EBV-positive cells, as well as tumour regression of NPC cell lines *in vivo*, in conjunction with radiotherapy. These results suggest that EBV-positive NPC can be effectively targeted by gene therapy strategies that exploit the oriP/EBNA1 relationship.

p53

p53 has been described as the 'guardian of the genome' and is the archetypal tumour suppressor gene; inducing cell cycle arrest and apoptosis in response to DNA damage. Low levels of p53 have been found in most head and neck cancers⁽⁵²⁾; however, NPC has been demonstrated to have p53 over-expression in between 31 - 95% of primary tumours⁽⁵³⁾. Mutations of p53 in NPC are rare, with the wild-type p53 induced apoptosis inactivated through loss of p14 and a mutated version of p63 (DN-p63) in NPC⁽⁵⁴⁾. p14 is essential in maintaining p53 stability⁽⁵⁵⁾. DN-p63 binds p53; thus preventing normal apoptosis activation⁽⁵⁶⁾. In spite of this, there is evidence of therapeutic efficacy of p53 gene therapy for NPC; *in vitro*, in animal models and, more recently, in humans. *In vitro* studies have focused on adenoviral mediated p53 (adv.p53) gene therapy on NPC cells compared to control virus;^(57, 58) in conjunction with radiotherapy^(59,60) or chemotherapy⁽⁶¹⁾. Increased cytotoxicity and apoptosis was demonstrated in NPC cells treated with p53 expressing adenovirus, independent of EBV status⁽⁵⁹⁾. In human NPC nude mice models, tumour regression was found in mice treated with adv.p53^(58,62). However, one study showed no improvement of adenovirus p53 treatment with radiotherapy compared to radiotherapy alone in nude mice bearing CNE-3 NPC xenografts⁽⁶³⁾. This was felt to be due to poor transduction of NPC cells with adenovirus (< 15%). Therefore despite no clear rationale for p53 gene therapy in the treatment of NPC, convincing anti-tumour efficacy of this strategy has been demonstrated.

p16

p16 is a regulator of the G1 phase of the cell cycle and was one of the first members of INK4 (inhibitor of cyclin-dependent kinase 4 – CDK4) family to be identified⁽⁶⁴⁾. Activation of various complexes of cyclins and cyclin-dependent kinases (CDK) permits the progression through cell cycle. p16, by competing for cyclin D binding with CDK4, results in G1 cell cycle arrest⁽⁶⁵⁾. Inactivation of p16 is a common event in NPC (60-80% of primary tumours) and is associated with a worse prognosis, reduced radiosensitivity and higher rates of recurrence^(66,67). Inactivation is achieved through multiple mechanisms including homozygous deletion, promoter hypermethylation and point mutation⁽⁶⁸⁾. Studies of other human p16 null cancers, including head and neck cancer, pancreas and lung⁽⁶⁹⁻⁷¹⁾, have

shown that adenoviral mediated gene transfer has resulted in tumour growth inhibition and apoptosis⁽⁷²⁾. It has been demonstrated *in vitro* that infection of a low p16 expressing NPC cell line (CNE-1) with a human p16 expressing replication-deficient adenovirus (adv.p16) resulted in significantly reduced cell survival; which was further reduced with the addition of radiotherapy. CNE-2Z NPC cell line has high endogenous p16 expression and showed significantly improved cell survival compared to CNE-1 with adv.p16 treatment⁽⁷²⁾. NPC xenografts using the same cell lines *in vivo*, with and without treatment with p16 expressing adenovirus in SCID mice, showed no tumour growth in the low p16 expressing cell line (CNE-1) treated with adv.p16; compared to sustained tumour growth in the high p16 expressing cell line (CNE-2Z) treated with adv. p16. Cytotoxicity of adv.p16 appears to be multi-mechanistic with evidence of G1 arrest, senescence, apoptosis and necrosis. Furthermore, another group has demonstrated that p16 restoration of a different p16 null NPC cell line, NPC/HK-1, using a plasmid containing the p16 gene under control of a CMV promoter, was able to induce cell growth inhibition and G1 phase cell cycle arrest⁽⁷³⁾. Therefore p16 corrective gene therapy in p16 deficient NPC would appear to be a promising therapeutic strategy.

Other strategies

Herpes simplex thymidine kinase cyto-reductive gene therapy has been investigated in NPC under the control of human telomerase reverse transcriptase (hTERT) promoter and CMV enhancer. hTERT expression is elevated in many nasopharyngeal tumours, and application of the non-toxic pro-drug ganciclovir converted to toxic ganciclovir triphosphate (catalysed by thymidine kinase) resulted in selective cytotoxicity to NPC cells *in vitro* and *in vivo*⁽⁷⁴⁾. Further strategies using RNA interference of oncogenes and other pro-cancer genes, such as angiogenesis mediators, have been described with promising results. Adenovirus mediated transfer of multiple short hairpin RNA (shRNA) to silence vascular endothelial growth factor (VEGF), hTERT, and Bcl-xL oncogene has been demonstrated to induce growth suppression and apoptosis in human NPC cell lines *in vitro* and *in vivo*⁽⁷⁵⁾.

Clinical trials of gene therapy for NPC

Recombinant replication-incompetent adenovirus with a human wild-type p53 (rAd-p53) replacing the E1 region has demonstrated safety and efficacy in clinical trials in patients with HNSCC^(36,37). rAd-p53 has also shown promising results alone or in conjunction with conventional treatments in other types of cancer, including lung, brain and bladder cancer⁽²³⁾. In 2003, the Chinese Food and Drug Administration approved rAd-p53 (trademarked as Gendicine by Shenzhen SiBiono GeneTech [SiBiono; Shenzhen, China]) for the treatment of head and neck cancer. As such Gendicine became the world's first

gene therapy product approved by a governmental agency for the treatment of cancer.

Recently, a randomised controlled clinical trial has evaluated rAd-p53 intratumoural injections in conjunction with radiotherapy compared to radiotherapy alone in patients with NPC with spread to regional lymph nodes⁽⁷⁶⁾. Tumours were biopsied before and after treatment; p53 protein, as well as downstream target proteins and vascular endothelial growth factor (VEGF) protein, were assessed using immunohistochemistry and reverse transcriptase polymerase chain reaction (RT-PCR). The tumour response to treatment was determined by computed tomography (CT). Complete response rates (disappearance of all target lesions) was 2.73 times higher in the rAd-p53 and radiotherapy group, compared to radiotherapy alone (66.7% compared to 24.4%). Five year locoregional failure rates were 2.7% for rAd-p53 and radiotherapy compared to 28% for radiotherapy alone. 7.5% and 11.7% higher five-year overall survival and disease-free survival rates were found with the combined treatment group respectively. Other than transient fever, no dose-limiting toxicity was encountered. Therefore rAd-p53 is a safe and efficacious treatment for NPC, as well as other head and neck cancers.

In 2005, H101 (Shanghai Sunway Biotech; Shanghai, China) a replication selective adenovirus became the world's first oncolytic virus product to be approved by a government agency for the treatment of NPC in combination with

cisplatin-based chemotherapy. The virus features the same E1B-55kDa deletion to the ONYX-015 virus, which renders viral replication confined to cancer cells, which, unlike non-cancer cells, are able to efficiently export late viral RNA in the absence of E1A-55kDa⁽⁷⁷⁾. A phase III randomised control trial compared intratumoural injection of H101 and cisplatin-based chemotherapy to chemotherapy alone. Overall response rates of 78.8% and 39.6% were recorded respectively, suggesting significant anti-tumour efficacy of the combined treatment⁽⁴⁰⁾.

Conclusion

Increased understanding of the molecular biology of cancer, as well as advances in techniques to engineer viruses with improved cancer selectivity and cancer killing properties, has established a government approved gene therapy product for the treatment of head and neck cancers, including NPC. Further research will develop treatments with improved anti-tumour potency, with clinical efficacy as monotherapies - without conjunctive contemporary modalities of radio- and chemo-therapy. Longer term goals of cancer treatment personalised to the individual patient's cancer genotype remains the ultimate aim of cancer gene therapy.

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References

- Parkin DM. Cancer incidence in five continents. Lyon: International Agency for Research on Cancer, 2002.
- Nielsen NH, Mikkelsen F, Hansen JP. Nasopharyngeal cancer in Greenland. The incidence in an Arctic Eskimo population. *Acta Pathol Microbiol Scand A*. 1977; 85: 850-858.
- Fandi A, Altun M, Azli N, Armand JP, Cvitkovic E. Nasopharyngeal cancer: epidemiology, staging, and treatment. *Semin Oncol*. 1994; 21: 382-397.
- Buell P. The effect of migration on the risk of nasopharyngeal cancer among Chinese. *Cancer Res*. 1974; 34: 1189-1191.
- Simons MJE, Shanmugaratnam KE. The biology of nasopharyngeal carcinoma. Geneva: International Union Against Cancer, 1982.
- Shanmugaratnam K, Sobin LH, WHO Collaborating Centre for the Histological Classification of Upper Respiratory Tract Tumours. Histological typing of upper respiratory tract tumours. Geneva: World Health Organization, 1978.
- Micheau C, Rilke F, Pilotti S. Proposal for a new histopathological classification of the carcinomas of the nasopharynx. *Tumori*. 1978; 64: 513-518.
- Chan AT. Head and neck cancer: treatment of nasopharyngeal cancer. *Ann Oncol* 2005; 16 Suppl 2: ii265-268.
- The Gde. Nasopharyngeal Carcinoma: etiology and control. Lyon: International agency for research on Cancer, 1978.
- Lee AW, Poon YF, Foo Wet al. Retrospective analysis of 5037 patients with nasopharyngeal carcinoma treated during 1976-1985: overall survival and patterns of failure. *Int J Radiat Oncol Biol Phys*. 1992; 23: 261-270.
- Kwong DL, Pow EH, Sham JS, et al. Intensity-modulated radiotherapy for early-stage nasopharyngeal carcinoma: a prospective study on disease control and preservation of salivary function. *Cancer*. 2004; 101: 1584-1593.
- McMillan AS, Pow EH, Kwong DL, et al. Preservation of quality of life after intensity-modulated radiotherapy for early-stage nasopharyngeal carcinoma: results of a prospective longitudinal study. *Head Neck*. 2006; 28: 712-722.
- Huncharek M, Kupelnick B. Combined chemoradiation versus radiation therapy alone in locally advanced nasopharyngeal carcinoma: results of a meta-analysis of 1,528 patients from six randomized trials. *Am J Clin Oncol*. 2002; 25: 219-223.
- Ma J, Mai HQ, Hong MH, et al. Results of a prospective randomized trial comparing neoadjuvant chemotherapy plus radiotherapy with radiotherapy alone in patients with locoregionally advanced nasopharyngeal carcinoma. *J Clin Oncol*. 2001; 19: 1350-1357.
- Preliminary results of a randomized trial comparing neoadjuvant chemotherapy (cisplatin, epirubicin, bleomycin) plus radiotherapy vs. radiotherapy alone in stage IV(> or = N2, M0) undifferentiated nasopharyngeal carcinoma: a positive effect on progression-free survival. International Nasopharynx Cancer Study Group. VUMCA I trial. *Int J Radiat Oncol Biol Phys*. 1996; 35: 463-469.
- Our inheritance, our future: the Department of Health's White Paper on genetics. London: NHS Confederation, 2003.
- Lehrman S. Virus treatment questioned after gene therapy death. *Nature*. 1999; 401: 517-518.
- Young LS, Searle PF, Onion D, Mautner V. Viral gene therapy strategies: from basic science to clinical application. *J Pathol*. 2006; 208: 299-318.

19. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011; 144: 646-674.
20. Edelstein ML, Abedi MR, Wixon J. Gene therapy clinical trials worldwide to 2007—an update. *J Gene Med*. 2007; 9: 833-842.
21. Vile RG, Russell SJ, Lemoine NR. Cancer gene therapy: hard lessons and new courses. *Gene Ther*. 2000; 7: 2-8.
22. Shiraishi K, Kato S, Han SY, et al. Isolation of temperature-sensitive p53 mutations from a comprehensive missense mutation library. *J Biol Chem*. 2004; 279: 348-355.
23. Peng Z. Current status of gene therapy in China: recombinant human Ad-p53 agent for treatment of cancers. *Hum Gene Ther*. 2005; 16: 1016-1027.
24. Mullen CA, Kilstrup M, Blaese RM. Transfer of the bacterial gene for cytosine deaminase to mammalian cells confers lethal sensitivity to 5-fluorocytosine: a negative selection system. *Proc Natl Acad Sci U S A*. 1992; 89: 33-37.
25. Lin X, Huang H, Li S, et al. A phase I clinical trial of an adenovirus-mediated endostatin gene (E10A) in patients with solid tumors. *Cancer Biol Ther*. 2007; 6: 648-653.
26. Yamamura M, Modlin RL, Ohmen JD, Moy RL. Local expression of antiinflammatory cytokines in cancer. *J Clin Invest*. 1993; 91: 1005-1010.
27. Lei N, Shen FB, Chang JH, et al. An oncolytic adenovirus expressing granulocyte macrophage colony-stimulating factor shows improved specificity and efficacy for treating human solid tumors. *Cancer Gene Ther*. 2009; 16: 33-43.
28. Kaufman HL, Rao JB, Irvine KR, Bronte V, Rosenberg SA, Restifo NP. Interleukin-10 enhances the therapeutic effectiveness of a recombinant poxvirus-based vaccine in an experimental murine tumor model. *J Immunother*. 1999; 22: 489-496.
29. Kirn DH, Thorne SH. Targeted and armed oncolytic poxviruses: a novel multi-mechanistic therapeutic class for cancer. *Nat Rev Cancer*. 2009; 9: 64-71.
30. Kaneda Y, Tabata Y. Non-viral vectors for cancer therapy. *Cancer Sci*. 2006; 97: 348-354.
31. Everts B, van der Poel HG. Replication-selective oncolytic viruses in the treatment of cancer. *Cancer Gene Ther*. 2005; 12: 141-161.
32. Shirakawa T. The current status of adenovirus-based cancer gene therapy. *Mol Cells*. 2008; 25: 462-466.
33. Volpers C, Kochanek S. Adenoviral vectors for gene transfer and therapy. *J Gene Med*. 2004; 6 Suppl 1: S164-171.
34. Vattemi E, Claudio PP. Adenoviral gene therapy in head and neck cancer. *Drug News Perspect*. 2006; 19: 329-337.
35. Raper SE, Chirmule N, Lee FS, et al. Fatal systemic inflammatory response syndrome in a ornithine transcarbamylase deficient patient following adenoviral gene transfer. *Mol Genet Metab*. 2003; 80: 148-158.
36. Clayman GL, el-Naggar AK, Lippman SM, et al. Adenovirus-mediated p53 gene transfer in patients with advanced recurrent head and neck squamous cell carcinoma. *J Clin Oncol*. 1998; 16: 2221-2232.
37. Clayman GL, Frank DK, Brusio PA, Goepfert H. Adenovirus-mediated wild-type p53 gene transfer as a surgical adjuvant in advanced head and neck cancers. *Clin Cancer Res*. 1999; 5: 1715-1722.
38. Nemunaitis J, Khuri F, Ganly I, et al. Phase II trial of intratumoral administration of ONYX-015, a replication-selective adenovirus, in patients with refractory head and neck cancer. *J Clin Oncol*. 2001; 19: 289-298.
39. Nemunaitis J, Ganly I, Khuri F, et al. Selective replication and oncolysis in p53 mutant tumors with ONYX-015, an E1B-55kD gene-deleted adenovirus, in patients with advanced head and neck cancer: a phase II trial. *Cancer Res*. 2000; 60: 6359-6366.
40. Xia ZJ, Chang JH, Zhang L, et al. [Phase III randomized clinical trial of intratumoral injection of E1B gene-deleted adenovirus (H101) combined with cisplatin-based chemotherapy in treating squamous cell cancer of head and neck or esophagus]. *Ai Zheng*. 2004; 23: 1666-1670.
41. Raab-Traub N, Flynn K, Pearson G, et al. The differentiated form of nasopharyngeal carcinoma contains Epstein-Barr virus DNA. *Int J Cancer*. 1987; 39: 25-29.
42. Yates JL, Guan N. Epstein-Barr virus-derived plasmids replicate only once per cell cycle and are not amplified after entry into cells. *J Virol*. 1991; 65: 483-488.
43. Wang L, Shan L, Lo KW, et al. Inhibition of nasopharyngeal carcinoma growth by RTA-expressing baculovirus vectors containing oriP. *J Gene Med*. 2008; 10: 1124-1133.
44. Goldsmith K, Bendell L, Frappier L. Identification of EBNA1 amino acid sequences required for the interaction of the functional elements of the Epstein-Barr virus latent origin of DNA replication. *J Virol*. 1993; 67: 3418-3426.
45. Judde JG, Spangler G, Magrath I, Bhatia K. Use of Epstein-Barr virus nuclear antigen-1 in targeted therapy of EBV-associated neoplasia. *Hum Gene Ther*. 1996; 7: 647-653.
46. Ragoczy T, Heston L, Miller G. The Epstein-Barr virus Rta protein activates lytic cycle genes and can disrupt latency in B lymphocytes. *J Virol*. 1998; 72: 7978-7984.
47. Zuo Y, Wu J, Xu Z, et al. Minicircle-oriP-IFN γ : a novel targeted gene therapeutic system for EBV positive human nasopharyngeal carcinoma. *PLoS One*. 2011; 6: e19407.
48. Li JH, Chia M, Shi W, et al. Tumor-targeted gene therapy for nasopharyngeal carcinoma. *Cancer Res*. 2002; 62: 171-178.
49. Wang Y, Xue SA, Hallden G, et al. Virus-associated RNA I-deleted adenovirus, a potential oncolytic agent targeting EBV-associated tumors. *Cancer Res*. 2005; 65: 1523-1531.
50. Yip KW, Li A, Li JH, et al. Potential utility of BimS as a novel apoptotic therapeutic molecule. *Mol Ther*. 2004; 10: 533-544.
51. Tsai ST, Fang SY, Jin YT, Su IJ, Yang BC. Analysis of the expression of Fas-L in nasopharyngeal carcinoma tissues. *Oral Oncol*. 1999; 35: 421-424.
52. Gasco M, Crook T. The p53 network in head and neck cancer. *Oral Oncol*. 2003; 39: 222-231.
53. Sheu LF, Chen A, Tseng HH, et al. Assessment of p53 expression in nasopharyngeal carcinoma. *Hum Pathol*. 1995; 26: 380-386.
54. Murono S, Yoshizaki T, Park CS, Furukawa M. Association of Epstein-Barr virus infection with p53 protein accumulation but not bcl-2 protein in nasopharyngeal carcinoma. *Histopathology*. 1999; 34: 432-438.
55. Zhang Y, Xiong Y, Yarbrough WG. ARF promotes MDM2 degradation and stabilizes p53: ARF-INK4a locus deletion impairs both the Rb and p53 tumor suppression pathways. *Cell*. 1998; 92: 725-734.
56. Crook T, Nicholls JM, Brooks L, O'Nions J, Allday MJ. High level expression of deltaN-p63: a mechanism for the inactivation of p53 in undifferentiated nasopharyngeal carcinoma (NPC)? *Oncogene*. 2000; 19: 3439-3444.
57. Li JH, Li P, Klamut H, Liu FF. Cytotoxic effects of Ad5CMV-p53 expression in two human nasopharyngeal carcinoma cell lines. *Clin Cancer Res*. 1997; 3: 507-514.
58. Zeng Y, Prabhu N, Meng R, Eldeiry W. Adenovirus-mediated p53 gene therapy in nasopharyngeal cancer. *Int J Oncol*. 1997; 11: 221-226.
59. Li JH, Huang D, Sun BF, et al. Efficacy of ionizing radiation combined with adenoviral p53 therapy in EBV-positive nasopharyngeal carcinoma. *Int J Cancer*. 2000; 87: 606-610.
60. Li JH, Lax SA, Kim J, Klamut H, Liu FF. The effects of combining ionizing radiation and adenoviral p53 therapy in nasopharyngeal carcinoma. *Int J Radiat Oncol Biol Phys*. 1999; 43: 607-616.
61. Weinrib L, Li JH, Donovan J, Huang D, Liu FF. Cisplatin chemotherapy plus adenoviral p53 gene therapy in EBV-positive and -negative nasopharyngeal carcinoma. *Cancer Gene Ther*. 2001; 8: 352-360.
62. Chen W, Lee Y, Wang H, et al. Suppression of human nasopharyngeal carcinoma cell growth in nude mice by the wild-type p53 gene. *J Cancer Res Clin Oncol*. 1992; 119: 46-48.
63. Lax SA, Chia MC, Busson P, Klamut HJ, Liu FF. Adenovirus-p53 gene therapy in human nasopharyngeal carcinoma xenografts. *Radiother Oncol*. 2001; 61: 309-312.
64. Serrano M, Hannon GJ, Beach D. A new regulatory motif in cell-cycle control causing specific inhibition of cyclin D/CDK4. *Nature*. 1993; 366: 704-707.
65. Serrano M. The tumor suppressor protein p16INK4a. *Exp Cell Res*. 1997; 237: 7-13.
66. Makitie AA, MacMillan C, Ho J, et al. Loss of p16 expression has prognostic significance in human nasopharyngeal carcinoma. *Clin Cancer Res*. 2003; 9: 2177-2184.
67. Hwang CF, Cho CL, Huang CC, et al. Loss of cyclin D1 and p16 expression correlates with local recurrence in nasopharyngeal carcinoma.

- cinoma following radiotherapy. *Ann Oncol.* 2002; 13: 1246-1251.
68. Lo KW, Huang DP. Genetic and epigenetic changes in nasopharyngeal carcinoma. *Semin Cancer Biol.* 2002; 12: 451-462.
69. Rocco JW, Li D, Liggett WH, Jr., et al. p16INK4A adenovirus-mediated gene therapy for human head and neck squamous cell cancer. *Clin Cancer Res.* 1998; 4: 1697-1704.
70. Naruse I, Heike Y, Hama S, Mori M, Saijo N. High concentrations of recombinant adenovirus expressing p16 gene induces apoptosis in lung cancer cell lines. *Anticancer Res.* 1998; 18: 4275-4282.
71. Kobayashi S, Shirasawa H, Sashiyama H, et al. P16INK4a expression adenovirus vector to suppress pancreas cancer cell proliferation. *Clin Cancer Res.* 1999; 5: 4182-4185.
72. Lee AW, Li JH, Shi W, et al. p16 gene therapy: a potentially efficacious modality for nasopharyngeal carcinoma. *Mol Cancer Ther.* 2003; 2: 961-969.
73. Wang GL, Lo KW, Tsang KS, et al. Inhibiting tumorigenic potential by restoration of p16 in nasopharyngeal carcinoma. *Br J Cancer.* 1999; 81: 1122-1126.
74. Shen CX, Wen Z, Qian YH, Mu SF, Guan XF. Targeted gene therapy of nasopharyngeal cancer in vitro and in vivo by enhanced thymidine kinase expression driven by human TERT promoter and CMV enhancer. *J Exp Clin Cancer Res.* 2010; 29:94.
75. Han JB, Tao ZZ, Chen SM, Kong YG, Xiao BK. Adenovirus-mediated transfer of tris-shRNAs induced apoptosis of nasopharyngeal carcinoma cell in vitro and in vivo. *Cancer Lett.* 2011; 309: 162-169.
76. Pan JJ, Zhang SW, Chen CB, et al. Effect of recombinant adenovirus-p53 combined with radiotherapy on long-term prognosis of advanced nasopharyngeal carcinoma. *J Clin Oncol.* 2009; 27: 799-804.
77. O'Shea CC, Johnson L, Bagus B, et al. Late viral RNA export, rather than p53 inactivation, determines ONYX-015 tumor selectivity. *Cancer Cell.* 2004; 6: 611-623.

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